Collagen VI-Related Myopathies: Genetic and Clinical Findings

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Abstract

Introduction: Collagen VI-related disorders are a group of heterogeneous muscular diseases due to mutations within the *COL6A1*, *COL6A2*, and *COL6A3* genes, encoding collagen VI as an essential component of the extracellular matrix. Here, we reported four patients affected by collagen VI-related disorders with genetic variants in *COL6A* genes.

Case Presentation: After a comprehensive clinical examination, four unrelated patients with muscular dystrophy were referred for genetic counseling. Whole-exome sequencing followed by *Insilco* analysis was done for one affected individual from each family. The analysis of genomic data revealed four different mutations within the *COL6A1*, *COL6A2*, and *COL6A3* genes in the affected individuals.

Conclusions: According to the previous reports, limb-girdle muscular dystrophy is inherited as autosomal dominant, and congenital myosclerosis phenotype is inherited in an autosomal recessive manner. Carrier testing and prenatal testing are possible if pathogenic variants are recognized in an affected family member.

Keywords: Collagen VI, Ullrich Congenital Muscular Dystrophy, *COL6A1*, *COL6A2*, *COL6A3*

1. Introduction

Inherited myopathies are inherited disorders known by progressive muscle weakness and are very similar and yet heterogeneous in their clinical, pathological, and genetic classifications. However, the maximum joint separation is formed based on the onset age and the concerned (e.g., facial, proximal, and distal) muscles (1, 2).

Although many inherited dystrophies are associated with the dysfunction of dystrophin and its related proteins, this is not a specific mechanism for inherited muscular dystrophy. A variety of cellular pathways are associated with disorders related to inherited muscle diseases (3). Examples are summarized in Table 1.

Breaking the link between the cytoskeleton and the extracellular matrix is understood to be a concern within the pathological process of a variety of transmissible muscle diseases, rare collagen VI-related disorders, autosomal recessive myosclerosis, Bethlem myopathy (BM), Ullrich congenital muscular dystrophy (UCMD), autosomal dominant limb-girdle muscular and dystrophy phenotype (4-7).

Inherited myopathies present in childhood with vari-

Table 1. The List of Gene Products Known to Associate with Muscular Dystrophies (3)

Gene product	Location	Phenotypes
Emerin	Nuclear envelope	X-EDMD
Lamin A/C	Nuclear envelope	AD-EDMD LGMD 1B
Dysferlin	Membrane	Distal myopathy LGMD 2B
Caveolin 3	Caveolae	LGMD 1C
Calpain 3	Cytoplasm	LGMD 2A
Integrin $\alpha 7$	Membrane	Congenital myopathy
Collagen VI	ECM	Bethlem myopathy
DMP	Neuromuscular junction	Myotonic dystrophy
ZNF9	Nuclear	PROMO
PABP2	Nuclear	OPMD
Telethonin	Sarcomeric	LGMD 2G
Myotilin	Sarcomeric	LGMD 1A

Abbreviations: EDMD, Emery-Dreifuss muscular dystrophy; LGMD, limb-girdle muscular dystrophy; DMPK, myotonic dystrophy protein kinase; PROMM, proximal myotonic myopathy; OPMD, oculopharyngeal muscular dystrophy; and PABP2, polyA-binding protein 2.

able symptoms such as hypotonia, feeding and metabolic process difficulties, and delayed motor skill development. Some patients with inherited less severe disease may present scoliosis, contraction, and respiratory failure later in life (1). Typically, clinical findings include general or proximal weakness, hypotension, muscle weakness, and secondary deformity due to muscle weakness. Usually, the diagnosis of inherited myopathies is made upon a detailed history and examination, followed by biochemical and neuroscience assessment and completed by muscle biopsy and genetic testing (5).

Although the involved muscles are sometimes concerned, the involvement of cardiac muscles is rare. Electromyography is ordinary and can show low-amplitude, polyphasic (myopathic) potentials. Changes specific to the myopathy are visible through muscle biopsy without fibrotic or necrotic changes (5, 8).

Bethlem and van Wijngaarden were the first to discover BM in 1976 (5). Normal flexion contractions, slowly progressive weakness of the proximal muscles, flexors of the long fingers, wrists, elbows, pectoralis major, and ankle dystrophy are characteristic of this myopathy. Patients with BM sometimes become symptomatic with hypotonia at birth. However, these patients will usually show symptoms in the first or second decade of life. Patients with BM continue walking until adulthood, but two-thirds of them need walking aid at the age of about sixty. The decline in respiratory function is variable and does not occur until adulthood in these patients (9). Noticeably, UCMD is a more severe form of the disease, with congenital weakness and tone, proximal joint contractions, and excessive loosening of the distal joints. Unlike BM, where respiratory involvement is relatively rare, patients with UCMD usually need respiratory support (10).

In UCMD, the onset is usually birth or during the preceding year of life with a delay in acquiring motor skills. Patients with UCMD never begin walking or, if they do, they will lose their ability to walk around the age of ten. A decline in pulmonary function occurs early, and thus beginning mechanical ventilation at night from the first or second decade of life is very important and helpful (11). The prevalence of BM and UCMD is reported to be 0.77:100,000 and 0.13:100,000, respectively, although this rate may not be precise as some patients could have been underdiagnosed (12). Here, we reported clinical and molecular genetic findings in four unrelated families.

2. Cases Presentation

2.1. Patient 1

The proband (P1) belonged to a 6-year-old symptomatic boy with myopathy. The patient's parents were relatives.

He had had a history of proximal muscle weakness since childhood. Creatine kinase levels mildly increased (about 540 U/L) in the patient, and his electromyography showed a myopathic pattern. There was history of affected individuals in their family with the same symptoms such as difficulty in walking, speech, and limb deformity.

2.2. Patient 2

The proband (P2) represented a 10-year-old boy. The patient's disease symptoms started at the age of five, including severe muscle atrophy and difficulty in walking and standing up. The patient's electromyography showed a myopathic pattern. Creatine kinase levels moderately increased (about 4000 U/L) in the patient constantly. In his examination, the muscular atrophy of the distal and proximal limbs was observed. The patient's father was also affected and had his son's exact symptoms.

2.3. Patient 3

The proband (P3) was a 17-year-old boy with high CK (2592 U/L) at the age of 14 who showed mild axial muscle weakness. He had trouble breathing, difficulty in standing up, fatigue, speech problems, and poor vision but had no heart problems. Creatine kinase levels moderately increased (about 2952 U/L) in the patient, and his electromyography showed irritable myopathy. The proband's 8-year-old sister presented with a similar clinical history and a milder phenotype: she could barely stand up and had difficulty breathing in her sleep. Also, one of her lower limbs was shorter than the other.

2.4. Patient 4

The proband (P4) was a 28-year-old male with creatine kinase levels of about 4708 U/L and normal echocardiography. The patient's muscle biopsy results showed that the striated muscles were involved in necrosis and atrophy in a non-fascicular manner. The proband walked with difficulty and the help of others. He also had progressive paralysis, and his sister and cousin had similar but milder symptoms.

The family pedigrees and main clinical manifestation of all the patients in the present study are shown in [Figure 1](#) and [Table 2](#), respectively.

3. Methods

We investigated many patients from different centers and referrals from neurologists to our Genetics Research Center. After genetic counseling, completing the relevant

Figure 1. The pedigrees of reported families. Black shapes show affected individuals. The chromatogram of validated variants through Sanger sequencing.

questionnaire, and obtaining consent form for the clinical diagnosis of myopathic diseases, all laboratory procedures, including muscle enzymes, creatine phosphokinase and aldolase, LDH, SGPT, SGOT, MRI, ECG, electromyography, and genetic testing, were briefly used. Five ml periph-

eral whole blood was collected in an EDTA tube from available family members who took part in genetic tests. The first step in genetic diagnostics is DNA extraction using the Gene All Kit. We performed whole exome sequencing (WES) through the Illumina HiSeq 4000 system (Illumina, Inc.,

Table 2. The Patients' Clinical Features

Patient	Age (y)	Onset (y)	sex	Max CK (U/L)	Pattern of Contractures	Severity
P1	6	< 2	Female	540	Proximal weakness	Mild
P2	10	> 4	Male	4000	Moderate proximal. Distal laxity	Moderate
P3	17	> 12	Male	2952	Moderate weakness	Moderate
P4	28	> 14	Male	4708	Proximal contractures, distal hyperextensibility	Severe

San Diego, CA, USA). FASTQ files were aligned to the human reference sequence (hg19) by the Burrows-Wheeler Aligner, and SAM files were produced.

Further, SAM to BAM conversion, BAM file sorting, and removal of duplicate reads were carried out using Picard (<http://picard.sourceforge.net>), followed by local realignment and variant calling using the Genome Analysis Tool Kit to generate VCF files. Then, annotation was performed with Annovar32 to identify pathogenic variants in affected individuals. Sanger sequencing was done using specific designed primers to confirm the candidate variants in patient and their parents.

3.1. Molecular Findings

The analysis results of *COL6A* genes in all patients included in the present study are summarized in Table 3. We detected a heterozygous variant in the exon9 of the *COL6A1* gene c.805G>A (p.Gly269Arg) in Patient 1. This variant has been described as the primary etiology for Ulrich's congenital muscular dystrophy (autosomal recessive and autosomal dominant). It is classified as a likely pathogenic variant (class 2) according to the recommendation of the American College of Medical Genetics and Genomics guideline. The parents and other family members should consider genetic counseling and confirmation of the variant by Sanger sequencing. Patient-4 revealed a homozygous mutation in the exon8 of the *COL6A1* gene c.784C>T (p.Arg262Trp) and showed the limb-girdle muscular dystrophy (LGMD) symptoms. One heterozygous mutation in the exon 15 of the *COL6A3* gene c.5581G>A (p.Glu1861Lys) was reported in Patient-2, although its frequency is very low in the average population.. BM 1 and Ulrich congenital muscular dystrophy 1 are inherited in an autosomal recessive/dominant manner. In Patient-3, a homozygous mutation was detected in the exon28 of the *COL6A2* gene (c.2917G>A, p.Val973Met). Also, a homozygous variant was detected in the exon8 of the *COL6A1* gene (c.784C>T, p.Arg262Trp) in Patient-4. This gene has been reported in association with autosomal recessive Ulrich's congenital muscular dystrophy1.

4. Discussion

Col-VI consists of three chains (alpha1, alpha2, and alpha3) encoded by totally different genes (*COL6A1*, *COL6A2*, and *COL6A3*). These three chains act within the cell and make a triple-helical compound assembled in dimers and tetramers stabilized by disulfide bonds. Each chain contains a relatively short triple helical domain with repetition Gly-X-Y subunits, flanked by sizeable globular N- and C-terminal domains (13, 14). The $\alpha 1$ (VI) and $\alpha 2$ (VI) chains are estimated to be around 1,000 amino acids long and have a molecular weight of 140-150 KDa. However, the $\alpha 3$ (VI) chain length is more significant as it contains numerous particles in the range of 2,500 to 3,100 amino acids and has a molecular weight of 250-300kDa (5, 15).

Collagen VI (Col-VI) is a protein extensively used in the extracellular matrix (ECM) of most connective tissues that help maintain tissue integrity and function (16). This protein is also considered a critical member of energy metabolism in adipose tissue, and low serum levels of this protein are reported related to fat distribution (15, 17). Col-VI is the main protein in the structure of multiple organs and tissues such as lungs, brain, myocardium, peripheral nerves, blood vessels, intervertebral disc, cornea, and skin (17).

Collagen VI-related disorders were first clinically recognized, and their pathophysiology was identified long after. These disorders have a range of different phenotypes, including BM, UCMD, limb-girdle muscular dystrophy, and myosclerosis myopathy. Some of these phenotypes may have overlapping clinical manifestations, but they have a different course of management, and this is a strong reason for specifying their clinical definitions (6, 7, 18).

According to muscle biopsy results, it is known that dystrophic features can be subcategorized to into degeneration, regeneration, and replacement of muscle tissue more often with fat tissue. Due to the genetic heterogeneity of the mentioned myopathies, the detection rate is relatively low. There have been some solutions, such as deletion/duplication analysis, but studies have shown no improvement in the detection rate. The coding region sequencing of the three genes (*COL6A1*, *COL6A2*, and *COL6A3*) has been accentuated by many reliable studies to confirm

Table 3. Molecular Findings Through WES Data

Patient	Gene	Ref	Nucleic Acid Alternation	Amino Acid Alternation	Exon	Heredity	Zygosity	Insilco Prediction
P1	COL6A1	NM_001848	c.805G>A	p. Gly269Arg	9	AR, AD	Het	likely pathogenic
P2	COL6A3	NM_057167	c.5581G>A	p. Glu1861Lys	15	AR, AD	Het	likely pathogenic
P3	COL6A2	NM_001849	c.2917G>A	p. Val973Met	28	AR, AD	Hom	VUS
P4	COL6A1	NM_001848	c.784C>T	p. Arg262Trp	8	AR, AD	Hom	VUS

Abbreviations: Het, heterozygous; Hom, homozygous; AR, autosomal recessive; AD, autosomal dominant; VUS, variant of unknown significance.

the detection of two mutant alleles (9, 19).

In conclusion, we found four different genetic variants in the COL6A1, COL6A2, and COL6A3 genes through a clinical genetic study. As a practical approach, NGS provides information for genetic counseling of related families. We hope that these findings help understand collagen VI-related myopathies.

Footnotes

Authors' Contribution: S.M. drafting of the manuscript, N.A. analysis and interpretation of data, M.D. analysis and interpretation of data, R.B. critical revision of the manuscript for important intellectual content, P.N.T. drafting of the manuscript, M.S. analysis and interpretation of data, M.F. drafting of the manuscript, M.R. drafting of the manuscript, S.A. drafting of the manuscript, H.M. drafting of the manuscript, E.G.H. study supervision.

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References

- Cardamone M, Darras BT, Ryan MM. Inherited myopathies and muscular dystrophies. *Semin Neurol*. 2008;**28**(2):250–9. doi: [10.1055/s-2008-1062269](#). [PubMed: [18351526](#)].
- Yates JR, Emery AE. A population study of adult onset limb-girdle muscular dystrophy. *J Med Genet*. 1985;**22**(4):250–7. doi: [10.1136/jmg.22.4.250](#). [PubMed: [4045950](#)]. [PubMed Central: [PMC1049444](#)].
- Burton EA, Davies KE. Muscular Dystrophy—Reason for Optimism? *Cell*. 2002;**108**(1):5–8. doi: [10.1016/S0092-8674\(01\)00626-2](#).
- Ullrich O. Kongenitale, atonisch-sklerotische Muskeldystrophie, ein weiterer Typus der hereditären degenerativen Erkrankungen des neuromuskulären Systems. *Zeitschrift für die gesamte Neurologie und Psychiatrie*. 1930;**126**(1):171–201. doi: [10.1007/bf02864097](#).
- Bethlem J, Wijngaarden GK. Benign myopathy, with autosomal dominant inheritance. A report on three pedigrees. *Brain*. 1976;**99**(1):91–100. doi: [10.1093/brain/99.1.91](#). [PubMed: [963533](#)].
- Scacheri PC, Gillanders EM, Subramony SH, Vedanarayanan V, Crowe CA, Thakore N, et al. Novel mutations in collagen VI genes: expansion of the Bethlem myopathy phenotype. *Neurology*. 2002;**58**(4):593–602. doi: [10.1212/wnl.58.4.593](#). [PubMed: [11865138](#)].
- Merlini L, Martoni E, Grumati P, Sabatelli P, Squarzonni S, Urciuolo A, et al. Autosomal recessive myosclerosis myopathy is a collagen VI disorder. *Neurology*. 2008;**71**(16):1245–53. doi: [10.1212/01.wnl.0000327611.01687.5e](#). [PubMed: [18852439](#)].
- Caria F, Cescon M, Gualandi F, Pichiecchio A, Rossi R, Rimessi P, et al. Autosomal recessive Bethlem myopathy: A clinical, genetic and functional study. *Neuromuscul Disord*. 2019;**29**(9):657–63. doi: [10.1016/j.nmd.2019.07.007](#). [PubMed: [31471117](#)].
- Lampe AK, Flanigan KM, Bushby KM, Hicks D. *Collagen type VI-related disorders*. University of Washington, Seattle, Seattle (WA): GeneReviews; 2012.
- Kirschner J, Bonnemant CG. The congenital and limb-girdle muscular dystrophies: sharpening the focus, blurring the boundaries. *Arch Neurol*. 2004;**61**(2):189–99. doi: [10.1001/archneur.61.2.189](#). [PubMed: [14967765](#)].
- Mercuri E, Yuva Y, Brown SC, Brockington M, Kinali M, Jungbluth H, et al. Collagen VI involvement in Ullrich syndrome: a clinical, genetic, and immunohistochemical study. *Neurology*. 2002;**58**(9):1354–9. doi: [10.1212/wnl.58.9.1354](#). [PubMed: [12011280](#)].
- Kirschner J. Congenital muscular dystrophies. *Handb Clin Neurol*. 2013;**113**:1377–85. doi: [10.1016/B978-0-444-59565-2.00008-3](#). [PubMed: [23622361](#)].
- Baldock C, Sherratt MJ, Shuttleworth C, Kieley CM. The Supramolecular Organization of Collagen VI Microfibrils. *Journal of Molecular Biology*. 2003;**330**(2):297–307. doi: [10.1016/S0022-2836\(03\)00585-0](#).
- Bonaldo P, Russo V, Bucciotti F, Bressan GM, Colombatti A. $\alpha 1$ Chain of Chick Type VI Collagen. *Journal of Biological Chemistry*. 1989;**264**(10):5575–80. doi: [10.1016/S0021-9258\(18\)83585-2](#).
- Rodriguez MA, Del Rio Barquero LM, Ortez CI, Jou C, Vigo M, Medina J, et al. Differences in Adipose Tissue and Lean Mass Distribution in Patients with Collagen VI Related Myopathies Are Associated with Disease Severity and Physical Ability. *Front Aging Neurosci*. 2017;**9**:268. doi: [10.3389/fnagi.2017.00268](#). [PubMed: [28848425](#)]. [PubMed Central: [PMC5550692](#)].
- Keene DR, Engvall E, Glanville RW. Ultrastructure of type VI collagen in human skin and cartilage suggests an anchoring function for this filamentous network. *J Cell Biol*. 1988;**107**(5):1995–2006. doi: [10.1083/jcb.107.5.1995](#). [PubMed: [3182942](#)]. [PubMed Central: [PMC2115316](#)].
- Engvall E, Hessel H, Klier G. Molecular assembly, secretion, and matrix deposition of type VI collagen. *J Cell Biol*. 1986;**102**(3):703–10. doi: [10.1083/jcb.102.3.703](#). [PubMed: [3456350](#)]. [PubMed Central: [PMC2114116](#)].
- Pepe G, Bertini E, Bonaldo P, Bushby K, Giusti B, de Visser M, et al. Bethlem myopathy (BETHLEM) and Ullrich scleroatonic muscular dystrophy: 100th ENMC International Workshop, 23–24 November 2001, Naarden, The Netherlands. *Neuromuscular Disorders*. 2002;**12**(10):984–93. doi: [10.1016/S0960-8966\(02\)00139-6](#).

19. Jimenez-Mallebrera C, Maioli MA, Kim J, Brown SC, Feng L, Lampe AK, et al. A comparative analysis of collagen VI production in muscle, skin and fibroblasts from 14 Ullrich congenital muscular dystrophy patients with dominant and recessive COL6A mutations. *Neuromuscul Disord.* 2006;**16**(9-10):571-82. doi: [10.1016/j.nmd.2006.07.015](https://doi.org/10.1016/j.nmd.2006.07.015). [PubMed: [16935502](https://pubmed.ncbi.nlm.nih.gov/16935502/)].