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Five-year Surveillance of Antimicrobial Resistance Changes and Epidemiological Characteristics in *Pseudomonas aeruginosa*: A Retrospective Study in a Chinese City Hospital

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Abstract

Background: In recent years, the widespread use of antibiotics has resulted in increased rates of antibiotic resistance (ABR). *Pseudomonas aeruginosa* is one of the most important opportunistic pathogens causing hospital-acquired infections. *Pseudomonas aeruginosa* has continuously increased resistance to commonly used clinical antimicrobial drugs, bringing great difficulties to clinical treatment.

Objectives: This retrospective study investigated the epidemiological characteristics of *P. aeruginosa* and changes in ABR over a 5-year period at a hospital in Shandong Province, China.

Methods: *Pseudomonas aeruginosa* strains were collected from 2015 to 2019. The antimicrobial susceptibility testing employed the Kirby-Bauer disk diffusion method and the broth microdilution method (VITEK-2 compact system), according to the guidelines by the Clinical and Laboratory Standards Institute. Data were analyzed using WHONET 5.6 and SPSS V. 21.0 software.

Results: A total of 3,324 *P. aeruginosa* strains were isolated from clinical specimens (604, 631, 700, 595, and 794 strains from 2015 to 2019, respectively). The highest *P. aeruginosa* detection rates were from respiratory tract specimens (72.54%). The highest resistance was seen in aztreonam, followed by ciprofloxacin, levofloxacin, and imipenem. The isolation rates for carbapenem-resistant *P. aeruginosa* (CRPA) and multidrug-resistant *P. aeruginosa* (MDRPA) ranged from 15.21-18.38% and 17.31-27.31%, respectively. Also, the isolation rates for extensively drug-resistant *P. aeruginosa* (XDRPA) ranged from 1.86-3.52%.

Conclusions: The main sources of the *P. aeruginosa* isolates were older adult patients with chronic respiratory diseases. The isolation rates for CRPA, MDRPA, and XDRPA strains decreased over the 5-year period. However, the drug resistance situation remains a serious concern. Hence, continued infection control and antimicrobial stewardship and basic and clinical research on bacterial resistance are essential.

Keywords: Pseudomonas aeruginosa, Antibiotic Resistance, Carbapenem-resistant Pseudomonas aeruginosa (CRPA), Multidrug-resistant Pseudomonas aeruginosa (MDRPA), Extensively Drug-resistant Pseudomonas aeruginosa (XDRPA)

1. Background

Pseudomonas aeruginosa is a non-fermenting Gramnegative bacillus and one of the most common bacteria causing nosocomial infections. Nosocomial infections by *P. aeruginosa* can cause blood diseases, respiratory diseases, urinary tract infections, immune disorders, and postoperative complications (1, 2). In recent years, the widespread use of antibiotics has resulted in increased rates of antibiotic resistance (ABR). In particular, the rates of ABR among *Acinetobacter baumannii* and *Klebsiella pneumoniae* isolates have increased continuously. Among numerous studies conducted on *P. aeruginosa*, two studies by Hu et al. have reported decreasing ABR trends in this

bacterium (1, 2). According to the National Healthcare Safety Network, carbapenem-resistant *P. aeruginosa* isolates occur more frequently than carbapenem-resistant *K. pneumonia* isolates. In most countries, the proportion of carbapenem-resistant *P. aeruginosa* is between 10% and 50% (3, 4). Unfortunately, the development of new drugs is far slower than the rate of ABR emergence, posing major challenges for the clinical treatment of bacterial infections and a significant threat to human health (4,5). ABR has become one of the largest public health threats of the 21st century. Therefore, monitoring of ABR is critically important.

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2. Objectives

Few studies have analyzed ABR of *P. aeruginosa* isolates in tandem with patients' clinical characteristics over a long follow-up period (3-5). Therefore, we investigated ABR among inpatient-derived *P. aeruginosa* isolates and the clinical characteristics of the affected patients in our hospital from 2015 to 2019. Our overall goal was to identify factors related to ABR and any changes in the epidemiology of *P. aeruginosa* over time.

3. Methods

3.1. Bacterial Isolates

The bacterial strains were collected from outpatients and inpatients from January 2015 to December 2019 at Weifang People's Hospital, a provincial regional medical center in Shandong Province, China. Only the first strain from each patient was included in this study. All strains were characterized using a VITEK-2 compact instrument (bioMérieux, France) according to the guidelines from the Clinical and Laboratory Standards Institute (CLSI) (6). All strains were isolated on Columbia blood plates (bioMérieux) prior to being characterized. Single colonies were used to make a 0.5 McFarland turbidity standard suspension. We obtained a written consent from all patients and the ethics committee of Weifang People's Hospital approved the study.

3.2. Antimicrobial Susceptibility Testing

For antimicrobial susceptibility testing, we used the Kirby-Bauer (K-B) disk diffusion method and broth microdilution method (VITEK-2 compact system), as recommended by the guidelines from the CLSI (6). Disk diffusion testing was performed on Muller-Hinton agar (Oxoid, UK) with the following disks: aztreonam (ATM), cefoperazone-sulbactam (CSL), piperacillin (PRL), and meropenem (MEM), according to the guidelines from the CLSI (6). Broth microdilution testing was performed on the VITEK-2 compact system to determine the minimum inhibitory concentrations (MICs) of the following antibiotics: ceftazidime (CAZ), ciprofloxacin (CIP), levofloxacin (LVX), cefepime (FEP), gentamicin (GEN), amikacin (AMK), tobramycin (TOB), piperacillin-tazobactam (TZP), and imipenem (IPM). Isolates shown to be resistant to IPM and/or MEM were defined as carbapenem-resistant P. aeruginosa (CRPA), and those resistant to three or more drugs class were defined as multidrug-resistant P. aeruginosa (MDRPA). Isolates with extensive drug resistance were defined as extensively drug-resistant P. aeruginosa (XDRPA). Pseudomonas aeruginosa ATCC 27853 and Escherichia coli ATCC25922 were used as controls for ABR.

3.3. Data Management and Statistical Analysis

Antimicrobial susceptibility testing data were centrally converted into a standard format using WHONET 5.6 (WHO, Switzerland), which was used for data management. SPSS 21.0 software (SPSS Inc., USA) was used for statistical analysis. Results were presented as numbers (percentages) for categorical variables, and the χ^2 test was used to compare them. Statistical significance was set at P < 0.05.

4. Results

4.1. Changes in the Pseudomonas aeruginosa Isolation Rate Over Time and Clinical Characteristics of the Patients

A total of 3,324 *P. aeruginosa* strains (excluding duplicates) were isolated from clinical specimens from 2015 to 2019, resulting in 604, 631, 700, 595, and 794 strains, respectively. The clinical characteristics of the patients with *P. aeruginosa* bacteremia are shown in Table 1. During this 5-year period, 65 - 67% of the cases were male. Older adults were the main affected population, among whom those aged 61 - 75 years had the highest percentage (30 - 41%), followed by those aged 46 to 60 (24 - 33%) and older than 75 (18 - 22%). Most isolates from the last five years were sourced from patients with *P. aeruginosa* pneumonia, the infection rate of which was 72.65% (range, 68.07 - 78.97%).

The second most common infection was soft tissue infection (infection rate, 18.32%; range, 13.08 - 22.52%). The third most common infection was urinary tract infection (infection rate, 3.61%; range, 2.69 - 4.97%). Blood flow-related infection was the fourth most common infection (rate, 3.28%; range, 2.15 - 4.53%). While other infections were less common, the incidence of intensive care unit (ICU)-related infections over the last five years was 11.85% (range, 10.71 - 14.07%). Over the last five years, the percentage of patients with malignancy-related diseases was 29.06% (range, 17.72 - 39.66%), those with cerebrovascular diseases was 22.68% (range, 18.38 - 27.15%), and those with pulmonary diseases was 22.65% (range, 17.12 - 27.81%).

4.2. Changing Trends in Antimicrobial Susceptibility

The antimicrobial susceptibility in *P. aeruginosa* is shown in Table 2. Except for the low value for GEN in 2018 from 1 to 0.5 μ g/mL, the MIC range for the clinical antibacterial drugs used over the past five years showed no changes. In contrast, the MIC₅₀ values for the antimicrobial drugs showed that LVX decreased from 1 to 0.5 μ g/mL in 2016 - 2018 and TZP from 8 to 4 μ g/mL after 2016, whereas the MIC₅₀ values of other drugs did not change. While the MIC₉₀ values of the clinical drugs CIP, FEP, and TZP did not change over the last five years, the values for GEN, TOB, and AMK aminoglycosides rose in 2016 and declined in 2017,

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Variables	2015 (N = 604)	2016 (N = 631)	2017 (N = 700)	2018 (N = 595)	2019 (N = 794)	Total (N = 3324)
Sex						
Male	407 (67.38)	417 (66.09)	463 (66.14)	394 (66.22)	520 (65.49)	2201 (66.22)
Female	197 (32.62)	214 (33.91)	237 (33.86)	201 (33.78)	274 (34.51)	1123 (33.78)
Age rang						
0 - 12	20 (3.31)	29 (4.6)	20 (2.86)	19 (3.19)	26 (3.27)	114 (3.43)
13 - 30	31 (5.13)	40 (6.34)	26 (3.71)	30 (5.04)	30 (3.78)	157 (4.72)
31 - 45	55 (9.11)	46 (7.29)	66 (9.43)	63 (10.59)	69 (8.69)	299 (9)
46-60	194 (32.12)	168 (26.62)	200 (28.57)	163 (27.39)	191 (24.06)	916 (27.56)
61 - 75	183 (30.3)	233 (36.93)	253 (36.14)	190 (31.93)	318 (40.05)	1177 (35.41)
> 75	121 (20.03)	115 (18.23)	135 (19.29)	130 (21.85)	160 (20.15)	661 (19.89)
ource of bacteremia						
Primary bacteremia	13 (2.15)	17 (2.69)	26 (3.71)	17 (2.86)	36 (4.53)	109 (3.28)
Pneumonia	477 (78.97)	474 (75.12)	494 (70.57)	405 (68.07)	565 (71.16)	2415 (72.65)
Urinary tract infection	30 (4.97)	17 (2.69)	21(3)	18 (3.03)	34 (4.28)	120 (3.61)
Intraabdominal infection	3 (0.5)	5 (0.79)	9 (1.29)	14 (2.35)	14 (1.76)	45 (1.35)
Soft tissue infection	79 (13.08)	110 (17.43)	146 (20.86)	134 (22.52)	140 (17.63)	609 (18.32)
Catheter related blood	2 (0.33)	6 (0.95)	2 (0.29)	6 (1.01)	4 (0.5)	20 (0.6)
Cerebrospinal fluid	0(0)	2 (0.32)	2 (0.29)	2 (0.34)	1(0.13)	7 (0.21)
ICU acquired	85 (14.07)	72 (11.41)	77 (11)	75 (12.61)	85 (10.71)	394 (11.85)
nderlying disease						
Malignancy	107 (17.72)	126 (19.97)	206 (29.43)	236 (39.66)	291 (36.65)	966 (29.06)
Hematologic malignancy	11 (1.82)	9 (1.43)	11 (1.57)	19 (3.19)	21 (2.64)	71 (2.14)
Solid tumor	96 (15.89)	117 (18.54)	195 (27.86)	217 (36.47)	270 (34.01)	895 (26.93)
Cerebral vascular stroke	164 (27.15)	116 (18.38)	144 (20.57)	161 (27.06)	169 (21.28)	754 (22.68)
Liver cirrhosis	3 (0.5)	4 (0.63)	2 (0.29)	5 (0.84)	7(0.88)	21 (0.63)
Diabetes mellitus	8 (1.32)	8 (1.27)	9 (1.29)	13 (2.18)	18 (2.27)	56 (1.68)
Cardiovascular diseases	37(6.13)	23 (3.65)	46 (6.57)	53 (8.91)	71 (8.94)	230 (6.92)
Chronic obstructive	31 (5.13)	28 (4.44)	42 (6)	61 (10.25)	81 (10.2)	243 (7.31)
Pulmonary disease	168 (27.81)	108 (17.12)	133 (19)	163 (27.39)	181 (22.8)	753 (22.65)
Burn	29 (4.8)	27 (4.28)	29 (4.14)	33 (5.55)	39 (4.91)	157 (4.72)
Digestive system	18 (2.98)	37 (5.86)	19 (2.71)	26 (4.37)	37 (4.66)	137 (4.12)
Urinary system	23 (3.81)	19 (3.01)	24 (3.43)	26 (4.37)	37 (4.66)	129 (3.88)
Orthopaedic diseases	13 (2.15)	32 (5.07)	27 (3.86)	41 (6.89)	39 (4.91)	152 (4.57)

^a Values are expressed as No. (%).

and IPM decreased in 2016 and increased during 2017-2018. The resistance rate (95% confidence interval [CI]) trends over the past five years, except for the expansion of IPM, ATM, and CSL in 2016-2017, showed a downwards trend for the other antibiotics, and the overall range had a left shift trend. As Figure 1 shows, over the past five years, almost all the ABR rates of the *P. aeruginosa* isolates were below 20%,

except for those of CIP and ATM in 2015 and 2018, which exceed 20%. The highest ABR rate for the *P. aeruginosa* isolates from our hospital was for ATM, followed by CIP, LVX, and IPM. While the mid-level resistance rates were for PRL, MEM, and CAZ, those for AMK, FEP, GEN, TOB, and TZP were all below 10%.

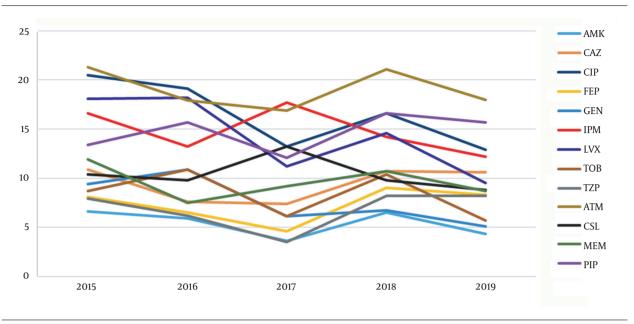


Figure 1. Trends in Pseudomonas aeruginosa antibiotic resistance rates from 2015 to 2019

4.3. Drug Sensitivity and Changes in CRPA, MDRPA, and XDRPA

By analyzing the clinical isolates for CRPA (Figure 2), we found that the drug resistance rates of the CRPA strains between 2015 and 2019 were 18.31%, 15.21%, 15.29%, 17.11%, and 15.74%, respectively. No statistical differences were seen for the values between 15% and 20%. The resistance rates for the MDRPA strains isolated between 2015 and 2019 were 25.99%, 27.31%, 21%, 17.31%, and 18.51%, respectively. The overall downwards trend, especially in 2018, was not statistically significant, even though it rose in 2019. The resistance rates for the XDRPA strains isolated between 2015 and 2019 were 15.73, 17.59%, 13%, 12.1%, and 10.2%, respectively (P > 0.05). After 2016, the trend decreased annually, indicating a statistically significant decrease.

5. Discussion

According to the US Centers for Disease Control and Prevention (CDC), *P. aeruginosa* accounted for 7.1% of hospital infections in 2011 (6, 7), a value not too dissimilar from that of the EPINE survey in Spain where *P. aeruginosa* was the second most common cause of hospital-acquired infections (10.5%) in 2016 (8). The increased deterioration of lung function seen in patients with chronic *P. aeruginosa* infections indicates that this bacterium causes high morbidity and mortality (9,10). By analyzing the infection characteristics of *P. aeruginosa* at our hospital over the past five years, we found that the isolates showed an increasing trend of ABR, which is consistent with the trends for

P. aeruginosa in China and other countries (11, 12). The characteristics of the infected patients showed that the proportion of male patients was larger than that of females, especially for elderly males over 60 years of age (ABR range, 48 - 63%), indicating that *P. aeruginosa* infection plays a very important role in infections in elderly males. This finding is consistent with other recent studies from around the world (13, 14).

Data from the isolates and patients over the last five years showed that infections caused by P. aeruginosa were mainly pneumonia (infection rate range, 68.07 - 78.97%). Soft tissue infections came next (infection rate range, 13.08 - 22.52%), followed by urinary tract infections (infection rate range, 2 - 4.97%) and blood flow-related infections (infection rate range, 2.15 - 4.53%). The incidence of ICU infections over the same period was 10.71 - 14.07%. Because P. aeruginosa is a conditional pathogenic bacterium, the normal respiratory tract and digestive tract, among other anatomical sites, can be colonized, especially in the elderly and in patients with poor immunity and susceptibility to opportunistic infections. Hence, the proportion of elderly patients with pulmonary infections was the highest in our hospital, a finding consistent with the data from China and abroad (1, 5). In this study, soft tissue infections were the second commonest source of isolates, a finding similar to the data of Li et al. (15), but different from the data by other studies (16-20). This discrepancy might be caused by the presence of *P. aeruginosa* in the natural environment, or by patients with skin and mucous membrane damage such as

burns from surgery, and other patients are also prone to nosocomial soft tissue infections.

The data from the *P. aeruginosa*-infected patients showed that the proportion with malignant tumors was nearly 30%, while those with cerebrovascular diseases and pulmonary diseases exceeded 20%. *Pseudomonas aeruginosa* infections were closely related to the immunity status and activity of the patients, and in this respect they are similar to the study by Huang et al. in the Taiwan Province of China (5). Thus, *P. aeruginosa* infections are mostly related to respiratory diseases, and this type of infection was at the forefront of ICU, a finding consistent with those of Huang et al. (5) and Feng et al. (21). Such infections are especially common in people with chronic diseases and in elderly patients may be related to many factors, such as decreased immunity in the elderly, long hospitalization times, and more interventional treatments (1, 21-23).

Table 1 shows that the MIC range of the clinical drugs did not change over the past five years. This indicates that, in terms of drug concentration, the antibiotic sensitivity range of P. aeruginosa was stable. The changing trend in MIC_{50} and MIC_{90} values was also mainly stable over time, except that the values for some of the drugs decreased. According to the changing trend for all the resistance rates (95% CI) across the past five years, the range had a downwards trend. Comparison of three values (resistance 95% CI, MIC_{50} , and MIC_{90}) and the average values for the hospital district from 2015 to 2019 showed that the change in drug resistance for all drug types had little fluctuations, and the overall drug resistance rate showed a downwards trend (5).

Drug sensitivity testing showed that the resistance rates were almost all lower than 20% during the study period, except that the drug resistance rates for CIP and ATM in 2015 and 2018 exceeded 20% against an overall decreasing trend. The drug resistance rate decreased significantly after 2016. The highest overall resistance rate was for ATM, followed by CIP and LVX, and the lowest drug resistance rate was for AMK, followed by TZP, and TOB. Our results showed that P. aeruginosa was more resistant to ATM and quinolones, but was more sensitive to PRL, TZP, and aminoglycosides, a finding mainly consistent with the monitoring data from CHINET reported by Hu et al. (1, 2). Another study (3) reported that the resistance rate of P. aeruginosa to AMK is low in Chongging and other parts of China, which is perhaps related to the substrate specificity of aminoglycoside-modifying enzyme-mediated drug resistance, and the frequency of using this drug by clinicians is also lower than that for other drugs. Because of the high nephrotoxicity of AMK, it is mainly used in combination with other antibiotics to treat P. aeruginosa infections to reduce possible side effects. In our previous study (24), we found that the most resistant genes in the variable region of the class I integrons are aminoglycoside resistance genes, which suggests that antibiotics should be used in combination to prevent drug resistance.

By analyzing changes in the resistance rate of P. aeruginosa to the same drugs across different years, we found that the drug resistance rate for ATM between 2016 and 2019 was significantly lower than that in 2015 (P < 0.05). The resistance rate for IPM was also lower than that in 2015, but there was no significant difference (P > 0.05). The remaining 10 drugs showed a trend of having the lowest resistance rates in 2019, which is consistent with previous studies (3-5, 15). This might be related to the antibiotic stewardship initiative campaign launched by the Chinese government in 2016 (3). It is generally believed that carbapenem resistance in P. aeruginosa is mainly related to the production of carbapenase and deletion or changes in the OprD2 outer membrane protein (14). It has been reported that the carbapenem resistance rate in *P. aeruginosa* is positively correlated with the use of such drugs.

The resistance rate for the P. aeruginosa clinical isolates was significantly lower than that for the unrestricted use of carbapenem antibiotics (25). In addition, some studies have shown that the primary bacteremia infection rate caused by XDRPA is higher than that caused by non-XDRPA (19). Although the XDRPA infection rate in our hospital was maintained below 5% in recent years, its continued monitoring is warranted. By analyzing the resistant patterns of P. aeruginosa, we found that CRPA strains in the past five years remained at 15 - 20%. MDRPA strains and XDRPA strains decreased from 27.31 to 18.51% and from 17.59% to 10.2%, respectively (Figure 2). Although the rates for these isolates were similar to those reported previously, the rates showed a statistically significant decrease, which did not correspond with those from other studies (25-30). Essentially, the trend appears to be a function of the Chinese government's antibiotic stewardship initiative campaign.

5.1. Conclusions

The main sources of *P. aeruginosa* isolates in this study were elderly patients with chronic respiratory diseases. *Pseudomonas aeruginosa* resistance rates for 12 commonly used antibiotics decreased over time. According to the obtained results, the isolation rate for CRPA, MDRPA, and XDRPA strains decreased over the past five years. It was a surprising discovery, while the drug resistance situation remains problematic. This study combined the ABR analysis and the pathogenic characteristics of *P. aeruginosa*. However, this was a single-center retrospective study. In the future, we plan to conduct a multicenter study to compare differences in ABR and the pathogenic characteristics of *P. aeruginosa*.

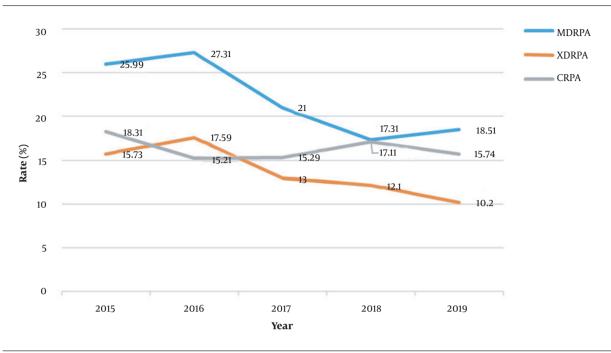


Figure 2. Resistance trends for CRPA, MDRPA, and XDRPA from 2015 to 2019

Footnotes

Authors' Contribution: Yansheng Ding designed and conducted the study, Yuhui Wang collected the data, Jie Ma and Wan-Xiang Li prepared the manuscript, Mi Liu provided advice and edited the manuscript. All authors have read and approved the final version of the manuscript.

Conflict of Interests: We declare that we have no conflicts of interest.

Data Reproducibility: The data presented in this study are openly available in one of the repositories or will be available on request from the corresponding author by this journal representative at any time during submission or after publication. Otherwise, all consequences of possible withdrawal or future retraction will be with the corresponding author.

Ethical Approval: The Ethics Committee of Weifang People's Hospital approved the study.

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Informed Consent: We obtained a written consent from all the patients.

References

- Hu F, Guo Y, Yang Y, Zheng Y, Wu S, Jiang X, et al. Resistance reported from China antimicrobial surveillance network (CHINET) in 2018. Eur J Clin Microbiol Infect Dis. 2019;38(12):2275–81. doi: 10.1007/s10096-019-03673-1. [PubMed: 31478103].
- Hu FP, Guo Y, Zhu D-M, Wang F, Jiang X-F, Xu Y-C, et al. Resistance trends among clinical isolates in China reported from CHINET surveillance of bacterial resistance, 2005–2014. Clin Microbiol Infect. 2016;22:9-14.
- 3. Feng W, Huang Q, Wang Y, Yuan Q, Li X, Xia P, et al. Changes in the resistance and epidemiological characteristics of Pseudomonas aeruginosa during a ten-year period. *J Microbiol Immunol Infect.* 2021. doi: 10.1016/j.jmii.2019.08.017. [PubMed: 31628088].
- Liu L, Liu B, Li Y, Zhang W. Successful control of resistance in Pseudomonas aeruginosa using antibiotic stewardship and infection control programs at a Chinese university hospital: a 6-year prospective study. *Infect Drug Resist.* 2018;11:637–46. doi: 10.2147/IDR.S163853. [PubMed: 29750044]. [PubMed Central: PMC5936004].
- Huang YC, Wu PF, Lin YT, Wang FD. Comparison of clinical characteristics of bacteremia from Elizabethkingia meningoseptica and other carbapenem-resistant, non-fermenting Gram-negative bacilli at a tertiary medical center. *J Microbiol Immunol Infect*. 2019;52(2):304–11. doi: 10.1016/j.jmii.2018.06.007. [PubMed: 30097388].
- Clinical and Laboratory Standards Institute. Performance Standards for Antimicrobial Susceptibility Testing; Twenty ninth informational supplement. Clin Lab Stand Ins. 2019.
- Katoh H, Yasumoto H, Shimizu M, Hamaoka S, Kinoshita M, Akiyama K. IV immunoglobulin for acute lung injury and bacteremia in Pseudomonas aeruginosa pneumonia. Crit Care Med. 2016;44. e12e24. doi: 10.1097/CCM.00000000000001271. [PubMed: 26317571].
- Ruiz-Garbajosa P, Cantón R. Epidemiology of antibiotic resistance in Pseudomonas aeruginosa. Implications for empiric and definitive therapy. Rev Esp Quimioter. 2017;30 (Suppl 1):8-12. [PubMed: 28882007].

- Lam JC, Somayaji R, Surette MG, Rabin HR, Parkins MD. Reduction in Pseudomonas aeruginosa sputum density during a cystic fibrosis pulmonary exacerbation does not predict clinical response.
 BMC Infect Dis. 2015;15:145. doi: 10.1186/s12879-015-0856-5. [PubMed: 25887462]. [PubMed Central: PMC4392784].
- Chin M, De Zoysa M, Slinger R, Gaudet E, Vandemheen KL, Chan F, et al. Acute effects of viral respiratory tract infections on sputum bacterial density during CF pulmonary exacerbations. *J Cyst Fibros*. 2015;14(4):482-9. doi: 10.1016/j.jcf.2014.11.009. [PubMed: 25544473]. [PubMed Central: PMC7105172].
- Djordjevic ZM, Folic MM, Jankovic SM. Previous antibiotic exposure and antimicrobial resistance patterns of Acinetobacter spp. and Pseudomonas aeruginosa isolated from patients with nosocomial infections. *Balkan Med J.* 2017;34(6):527-33. doi: 10.4274/balkanmedj.2016.1844. [PubMed: 29215335]. [PubMed Central: PMC5785657].
- Djordjevic Z, Folic MM, Zivic Z, Markovic V, Jankovic SM. Nosocomial urinary tract infections caused by Pseudomonas aeruginosa and Acinetobacter species: sensitivity to antibiotics and risk factors.
 Am J Infect Control. 2013;41(12):1182-7. doi: 10.1016/j.ajic.2013.02.018. [PubMed: 23726656].
- Aloush V, Navon-Venezia S, Seigman-Igra Y, Cabili S, Carmeli Y. Multidrug-resistant Pseudomonas aeruginosa: risk factors and clinical impact. *Antimicrob Agents Chemother*. 2006;50(1):43–8. doi: 10.1128/AAC.50.1.43-48.2006. [PubMed: 16377665]. [PubMed Central: PMC1346794].
- Zavascki AP, Carvalhaes CG, Picao RC, Gales AC. Multidrug-resistant Pseudomonas aeruginosa and Acinetobacter baumannii: resistance mechanisms and implications for therapy. Expert Rev Anti Infect Ther. 2010;8(1):71-93. doi: 10.1586/eri.09.108. [PubMed: 20014903].
- Li L, Dai JX, Xu L, Chen ZH, Li XY, Liu M, et al. Antimicrobial resistance and pathogen distribution in hospitalized burn patients: A multicenter study in Southeast China. *Medicine (Baltimore)*. 2018;97(34). e11977. doi: 10.1097/MD.0000000000011977. [PubMed: 30142829]. [PubMed Central: PMC6113017].
- European Centre for Disease Prevention and Control. Antimicrobial resistance surveillance in Europe 2015: Annual Report of the European Antimicrobial Resistance Surveillance Network (EARS-Net). Stockholm. 2017.
- 17. Cabot G, Ocampo-Sosa AA, Tubau F, Macia MD, Rodriguez C, Moya B, et al. Overexpression of AmpC and efflux pumps in Pseudomonas aeruginosa isolates from bloodstream infections: prevalence and impact on resistance in a Spanish multicenter study. *Antimicrob Agents Chemother*. 2011;**55**(5):1906–11. doi: 10.1128/AAC.01645-10. [PubMed: 21357294]. [PubMed Central: PMC3088238].
- Sader HS, Farrell DJ, Flamm RK, Jones RN. Antimicrobial susceptibility of Gram-negative organisms isolated from patients hospitalised with pneumonia in US and European hospitals: results from the SENTRY Antimicrobial Surveillance Program, 2009-2012. *Int J Antimicrob Agents*. 2014;43(4):328-34. doi: 10.1016/j.ijantimicag.2014.01.007. [PubMed: 24630306].
- Montero M, Sala M, Riu M, Belvis F, Salvado M, Grau S, et al. Risk factors for multidrug-resistant Pseudomonas aeruginosa acquisition. Impact of antibiotic use in a double case-control study. Eur J Clin Microbiol Infect Dis. 2010;29(3):335–9. doi: 10.1007/s10096-009-0850-1. [PubMed: 20033749].

- Fernandez-Barat L, Ferrer M, De Rosa F, Gabarrus A, Esperatti M, Terraneo S, et al. Intensive care unit-acquired pneumonia due to Pseudomonas aeruginosa with and without multidrug resistance. *J Infect*. 2017;74(2):142–52. doi: 10.1016/j.jinf.2016.11.008. [PubMed: 27865895].
- 21. Feng W, Sun F, Wang Q, Xiong W, Qiu X, Dai X, et al. Epidemiology and resistance characteristics of Pseudomonas aeruginosa isolates from the respiratory department of a hospital in China. *J Glob Antimicrob Resist.* 2017;**8**:142–7. doi: 10.1016/j.jgar.2016.11.012. [PubMed: 28216097].
- Haeili M, Ghodousi A, Nomanpour B, Omrani M, Feizabadi MM. Drug resistance patterns of bacteria isolated from patients with nosocomial pneumonia at Tehran hospitals during 2009-2011. J Infect Dev Ctries. 2013;7(4):312-7. doi:10.3855/jidc.2604. [PubMed: 23592640].
- Yayan J, Ghebremedhin B, Rasche K. Antibiotic resistance of Pseudomonas aeruginosa in pneumonia at a single university hospital center in Germany over a 10-year period. *PLoS One*. 2015;10(10). e0139836. doi: 10.1371/journal.pone.0139836. [PubMed: 26430738]. [PubMed Central: PMC4592231].
- Liu M, Ma J, Jia W, Li W. Antimicrobial resistance and molecular characterization of gene cassettes from class 1 integrons in Pseudomonas aeruginosa strains. *Microb Drug Resist.* 2020;26(6):670-6. doi: 10.1089/mdr.2019.0406. [PubMed: 32407190]. [PubMed Central: PMC7307683].
- Pakyz AI, Oinonen M, Polk RE. Relationship of carbapenem restriction in 22 university teaching hospitals to carbapenem use and carbapenem-resistant Pseudomonas aeruginosa. *Antimicrob Agents Chemother*. 2009;53(5):1983-6. doi: 10.1128/AAC.01535-08. [PubMed: 19273670]. [PubMed Central: PMC2681502].
- Horcajada JP, Montero M, Oliver A, Sorli L, Luque S, Gomez-Zorrilla S, et al. Epidemiology and treatment of multidrug-resistant and extensively drug-resistant Pseudomonas aeruginosa infections. Clin Microbiol Rev. 2019;32(4). doi: 10.1128/CMR.00031-19. [PubMed: 31462403]. [PubMed Central: PMC6730496].
- 27. Pena C, Cabot G, Gomez-Zorrilla S, Zamorano L, Ocampo-Sosa A, Murillas J, et al. Influence of virulence genotype and resistance profile in the mortality of Pseudomonas aeruginosa bloodstream infections. *Clin Infect Dis.* 2015;**60**(4):539–48. doi: 10.1093/cid/ciu866. [PubMed: 25378459].
- 28. Walkty A, Lagace-Wiens P, Adam H, Baxter M, Karlowsky J, Mulvey MR, et al. Antimicrobial susceptibility of 2906 Pseudomonas aeruginosa clinical isolates obtained from patients in Canadian hospitals over a period of 8 years: Results of the Canadian Ward surveillance study (CANWARD), 2008-2015. Diagn Microbiol Infect Dis. 2017;87(1):60-3. doi: 10.1016/j.diagnicrobio.2016.10.003. [PubMed: 28336136]
- Sader HS, Castanheira M, Duncan LR, Flamm RK. Antimicrobial susceptibility of Enterobacteriaceae and Pseudomonas aeruginosa isolates from United States medical centers stratified by infection type: results from the international network for optimal resistance monitoring (INFORM) surveillance program, 2015-2016. *Diagn Microbiol Infect Dis*. 2018;92(1):69–74. doi: 10.1016/j.diagmicrobio.2018.04.012. [PubMed: 29789189].
- Magill SS, Edwards JR, Bamberg W, Beldavs ZG, Dumyati G, Kainer MA, et al. Multistate point-prevalence survey of health care-associated infections. N Engl J Med. 2014;370(13):1198–208. doi: 10.1056/NEJ-Moa1306801. [PubMed: 24670166]. [PubMed Central: PMC4648343].

Table 2. Changes in Antimicrobial Susceptibility Trends from 2015 to 2019

Agent and Year	Resistance (R%)	Medium (R%)	Sensitive (R%)	Resistance Rate (95% CI)	MIC ₅₀	MIC90	District Average	MIC Range	P-Value		
VITEK-2 Compact Broth Micro-dilution Method											
AMK									0.114		
2015	6.6	1.5	91.9	4.3 - 10.0	2	8	3.179	2-64			
2016	5.9	0.6	93.5	3.7 - 9.1	2	16	3.145	2 - 64			
2017	3.6	0.8	95.7	2.1 - 6.1	2	8	2.964	2-64			
2018	6.5	0	93.5	4.3 - 9.7	2	8	2.967	2 - 64			
2019	4.3	0.6	95.2	2.5 - 7.1	2	8	3.064	2-64			
CAZ		-							0.045		
2015	10.9	6.7	82.4	7.8 - 14.9	4	32	4.269	1-64			
2016	7.6	10.9	81.5	5.1 - 11.1	4	16	4.689	1-64			
2017	7.4	11.5	81.1	5.1-10.6	4	16	4.616	1-64			
2018	10.7	5.3	84	7.8 - 14.5	4	32	4.888	1-64			
2019	10.6	8.9	80.5	7.7 - 14.4	4	32	5.178	1-64			
CIP									0.000		
2015	20.5	5.1	74.3	16.4 - 25.3	0.25	4	0.59	.25 - 4			
2016	19.1	7.9	72.9	15.1 - 23.8	0.25	4	0.596	.25 - 4			
2017	13.2	6.4	80.4	10.1-17.0	0.25	4	0.484	.25 - 4			
2018	16.6	3.9	79.4	13.0 - 21.0	0.25	4	0.485	.25 - 4			
2019	12.9	4.6	82.5	9.7-17.0	0.25	4	0.455	.25 - 4			
EP									0.012		
2015	8.1	5.4	86.4	5.5 - 11.7	2	16	2.918	1-64			
2016	6.5	6.8	86.8	4.2 - 9.8	2	16	3.236	1-64			
2017	4.6	6.6	88.8	2.8 - 7.3	2	16	3.004	1-64			
2018	9	3.9	87.1	6.3 - 12.6	2	16	3.28	1-64			
2019	8.3	4.3	87.4	5.7 - 11.8	2	16	3.23	1-64			
GEN									0.000		
2015	9.4	1.8	88.8	6.6 - 13.2	1	8	1.443	1-16			
2016	10.9	3.2	85.9	7.9 - 14.8	1	16	1.638	1-16			
2017	6.1	2.8	91	4.0 - 9.1	1	4	1.536	1-16			
2018	6.7	7.3	86	4.4 - 9.9	1	8	1.568	.5 - 16			
2019	5.1	3.7	91.2	3.1 - 8.1	1	4	1.556	1-16			
PM									0.017		
2015	16.6	6.9	76.5	12.8 - 21.1	2	16	2.364	1-16			
2016	13.2	10.6	76.2	9.9 - 17.4	2	8	2.179	1-16			
2017	17.7	1.8	80.5	13.5 - 22.8	2	16	2.57	1-16			
2018	14.2	0.4	85.4	10.4 - 19.0	2	16	2.308	1-16			
2019	12.2	0.8	87.1	8.6 - 16.9	2	8	2.148	1-16			
.VX									0.000		
2015	18.1	5.7	76.2	14.2 - 22.8	1	8	1.119	.25 - 8			
2016	18.2	5.3	76.5	14.3 - 22.8	0.5	8	1.101	.25 - 8			
2017	11.2	5.6	83.2	8.3 - 14.8	0.5	8	0.938	.25 - 8			
2018	14.6	4.2	81.2	11.2 - 18.8	0.5	8	0.907	.25 - 8			
2019	9.5	4.9	85.7	6.7 - 13.2	1	4	0.913	.25 - 8			
ОВ									0.00		
2015	8.7	0.9	90.4	6.0 - 12.4	1	4	1.342	1-16			
2016	10.9	1.2	87.9	7.9 - 14.8	1	16	1.435	1-16			
2017	6.1	0.5	93.4	4.0 - 9.1	1	1	1.225	1-16			
2018	10.4	0.8	88.8	7.5 - 14.2	1	16	1.392	1-16			
2019	5.7	1.7	92.6	3.6 - 8.8	1	2	1.25	1-16			
ZP									0.002		
2015	7.9	14.5	77.6	5.3 - II.5	8	64	9.767	4 - 128			
2016	6.2	16.9	76.9	4.0 - 9.5	4	64	9.484	4 - 128			
2017	3.5	13.9	82.6	2.0 - 6.1	4	64	8.198	4 - 128			

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	2018	8.2	7.9	84	5.5 - 11.9	4	64	8.266	4 - 128	
	2019	8.2	5.5	86.3	5.4 - 12.1	4	64	7.795	4 - 128	
K-B Method										
ATM										0.130
	2015	21.3	19.1	59.6	17.3 - 25.9					
	2016	17.9	13.9	68.2	12.3 - 25.2					
	2017	16.9	17.1	66	13.6 - 20.8					
	2018	21.1	13.2	65.8	17.3 - 25.5					
	2019	18	14.4	67.5	14.4 - 22.3					
CSL										0.081
	2015	10.4	17.3	72.3	7.5 - 14.1					
	2016	9.8	13.4	76.8	7.1 - 13.3					
	2017	13.2	13.2	73.5	8.4 - 19.9					
	2018	9.8	15.5	74.7	7.1 - 13.3					
	2019	8.8	14	77.2	6.2 - 12.3					
MEM										0.061
	2015	11.9	2.4	85.8	8.7 - 16.0					
	2016	7.5	2.3	90.2	5.2 - 10.7					
	2017	9.2	1.8	89	6.8 - 12.4					
	2018	10.7	2.5	86.8	7.9 - 14.2					
	2019	8.7	1.5	89.8	6.2 - 12.0					
PIP										0.116
	2015	13.4	0	86.6	10.2 - 17.4					
	2016	15.7	0	84.3	12.4 - 19.7					
	2017	12.1	0	87.9	9.3 - 15.6					
	2018	16.6	0	83.4	13.2 - 20.7					
	2019	15.7	0	84.3	12.3 - 19.7					

Abbreviation: CI, confidence interval.